INVITED REVIEW

Beyond the horizon: Innovations and future directions in axial-spondyloarthritis

Vincenzo Venerito¹*^(D), Sergio Del Vescovo¹*^(D), Giuseppe Lopalco¹^(D), Fabian Proft²^(D)

¹Department of Precision and Regenerative Medicine and Ionian Area (DiMePRe-J), Polyclinic Hospital, University of Bari, Bari, Italy ²Department of Gastroenterology, Infectiology and Rheumatology (including Nutrition Medicine), Charité-Universitätsmedizin Berlin, corporate member of Freie Universität Berlin and Humboldt-Universität zu Berlin, Berlin, Germany

Correspondence: Fabian Proft, MD.

E-mail: Fabian.Proft@charite.de

* These authors contributed equally to this study.

Received: November 18, 2023 Accepted: November 18, 2023 Published online: November 22, 2023

Citation: Venerito V, Vescovo SD, Lopalco G, Proft F. Beyond the horizon: Innovations and future directions in axial-spondyloarthritis. Arch Rheumatol 2023;38(4):491-511. doi: 10.46497/ ArchRheumatol.2023.10580.

This is an open access article under the terms of the Creative Commons Attribution-NonCommercial License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited and is not used for commercial purposes (http://creativecommons.org/ Licenses/by-nc/4.0/).

ABSTRACT

Axial spondyloarthritis (axSpA) is a chronic inflammatory disease of the spine and sacroiliac joints. This review discusses recent advances across multiple scientific fields that promise to transform axSpA management. Traditionally, axSpA was considered an immune-mediated disease driven by human leukocyte antigen B27 (HLA-B27), interleukin (IL)-23/IL-17 signaling, biomechanics, and dysbiosis. Diagnosis relies on clinical features, laboratory tests, and imaging, particularly magnetic resonance imaging (MRI) nowadays. Management includes exercise, lifestyle changes, non-steroidal anti-inflammatory drugs and if this is not sufficient to achieve disease control also biological and targeted-synthetic disease modifying anti-rheumatic drugs. Beyond long-recognized genetic risks like HLA-B27, high-throughput sequencing has revealed intricate gene-environment interactions influencing dysbiosis, immune dysfunction, and aberrant bone remodeling. Elucidating these mechanisms promises screening approaches to enable early intervention. Advanced imaging is revolutionizing the assessment of axSpA's hallmark: sacroiliac bone-marrow edema indicating inflammation. Novel magnetic resonance imaging (MRI) techniques sensitively quantify disease activity, while machine learning automates complex analysis to improve diagnostic accuracy and monitoring. Hybrid imaging like synthetic MRI/computed tomography (CT) visualizes structural damage with new clarity. Meanwhile, microbiome analysis has uncovered gut ecosystem alterations that may initiate joint inflammation through HLA-B27 misfolding or immune subversion. Correcting dysbiosis represents an enticing treatment target. Moving forward, emerging techniques must augment patient care. Incorporating patient perspectives will be key to ensure innovations like genetics, microbiome, and imaging biomarkers translate into improved mobility, reduced pain, and increased quality of life. By integrating cutting-edge, multidisciplinary science with patients' lived experience, researchers can unlock the full potential of new technologies to deliver transformative outcomes. The future is bright for precision diagnosis, tightly controlled treatment, and even prevention of axSpA.

Keywords: Artificial intelligence, axial spondyloarthritis, future directions, imaging, innate immunity.

Spondyloarthritis refers to a group of chronic inflammatory diseases that share common clinical features including inflammatory back pain, peripheral arthritis, enthesitis, uveitis, psoriasis, and inflammatory bowel disease (IBD). This disease concept encompasses several interrelated but distinct disorders: axial spondyloarthritis (axSpA), psoriatic arthritis (PsA), reactive arthritis (ReA), IBD-associated arthritis, and undifferentiated spondyloarthritis (Figure 1).¹ They each possess distinct clinical phenotypes and patterns of joint involvement. AxSpA primarily affects the spine and sacroiliac joints (SIJ); PsA manifests mainly as peripheral-often asymmetric-oligoarthritis, axial disease, enthesitis, and dactylitis; ReA classically follows certain infections and presents with lower extremity arthritis and enthesitis; IBD-associated spondyloarthritis occurs in patients with IBDs like Crohn disease and ulcerative colitis.¹

Specifically, axSpA refers to a group of chronic inflammatory diseases primarily affecting the SIJ and spine.² AxSpA commonly includes two clinical pictures: non-radiographic axSpA (nr-axSpA) and radiographic axSpA (r-axSpA), historically termed ankylosing spondylitis (AS), depending on whether the axial disease has caused visible radiographical lesions on X-ray (fulfilling the modified New York Criteria (NYc) or not).³ On the contrary, the



Figure 1. Spondyloarthritis spectrum from axial to peripheral involvement. SpA: Spondyloarthritis; IBD: Inflammatory bowel disease. Modified from Proft F et al. Ther Adv Musculoskelet Dis 2018;10:129-39.

notion of nr-axSpA frames the early phases of the disease, in which the axial involvement is marked by the presence of bone-marrow edema (BME) in the SIJ visible on magnetic resonance imaging (MRI). Therefore, these two entities essentially represent a continuum between an early and a more advanced stage of the same disease.⁴

The global prevalence of axSpA is estimated between 0.1-1.4%, with considerable geographic variation mainly attributed to differences in human leukocyte antigen B27 (HLA-B27) prevalence. HLA-B27, present in 8-10% of the general population, is positive in up to 90% of axSpA patients and is a major genetic risk factor. Prevalence is highest in Arctic and Northern European regions where HLA-B27 rates approach 50%, such as among the Haida native peoples of Western Canada with axSpA prevalence from 6-10%. In contrast, prevalence is markedly lower in Japan and Arab populations where HLA-B27 rates are only $1-3\%^{5-7}$. On the population level, prevalence estimates range from 0.24% in Greece to 1.8% in Northern Norway.^{8,9} A systematic review reported mean axSpA prevalence in Europe of 24 per 10,000 people. In Asia the estimate was 17 per 10,000, while North America ranged from 13 to 32 per 10,000.¹⁰ Although less studied than prevalence, incidence rates ranged from 5 to 15 per 100,000 person-years across studied populations.¹¹

The onset of axSpA symptoms usually begins in early adulthood, resulting in a substantial lifetime burden. Patients suffer from chronic back pain typically with inflammatory characteristics, spinal stiffness, and reduced mobility.¹² Therefore, the disease is also associated with major economic implications stemming from direct medical costs and indirect costs due to lost work productivity. Furthermore, structural damage is irreversible and leads to increasing disability, loss of quality of life, and need for surgical interventions over time.¹³⁻¹⁵

Despite the availability of effective treatments, an unsolved problem remains the average delay from symptom onset to diagnosis, which remains around 5-10 years globally.^{16,17} This diagnostic delay has motivated intensive research to better understand the pathophysiology and natural history of axSpA, and to optimize strategies for early identification and treatment.¹⁸ The emergence of MRI has been revolutionary, allowing direct visualization of SIJ inflammation years before radiographic changes appear and with this giving the opportunity for early and intensified treatments by suppressing the inflammatory activity enabling to even preventing the development of such structural changes. Therefore, incorporation of MRI findings into classification criteria has enabled earlier diagnosis and treatment.¹⁹

The discovery of tumor-necrosis factoralpha (TNF- α) and interleukin (IL)-17 as major inflammatory cytokines in axSpA led to the rise of biologic drugs targeting these mediators. Biologics have drastically advanced management of axSpA, though significant unmet needs remain.²⁰ Treatment responses vary widely and many questions persist surrounding their longterm impacts on controlling symptoms, preserve function, prevent the development of structural damage, and extra-articular manifestations. There is a need for prognostic biomarkers to predict disease course and response to therapies.

While advances in MRI imaging and biologic therapies have transformed treatment for many patients with axSpA critical gaps remain in reducing the years-long diagnostic delays that many patients face. To address this, optimizing screening and referral strategies in primary care is crucial, particularly for at-risk individuals with chronic back pain. Incorporating clinical, laboratory, genetic, and imaging biomarkers into predictive models is a promising approach to quantify axSpA risk earlier and enable prompt diagnosis and treatment, with the goal of improving long-term outcomes. Looking ahead, further research on disease mechanisms and new collaborations across disciplines will be key to advancing the field of axSpA and reducing the associated burden of the disease. Emerging fields include genetics to identify new risk factors, immunology to elucidate pathogenic pathways, microbiome studies to understand links with gut dysbiosis, advanced imaging techniques to improve diagnosis and monitoring, and digital tools for automated analysis and personalized medicine.

In summary, leveraging new technologies in genetics, immunology, microbiome research, imaging, and artificial intelligence (AI) holds tremendous potential to uncover axSpA disease insights, enable precision diagnosis and treatment, and ultimately improve the lives of patients struggling with this challenging disease. By pursuing a multidisciplinary approach across these exciting fields, researchers can pave the way for the next generation of innovations in axSpA care.²¹

TRADITIONAL UNDERSTANDINGS

Pathophysiology

Axial spondyloarthritis results from multiple interacting factors including genetic risks, immune dysregulation, biomechanics, and environmental triggers.²² While PsA is traditionally considered an enthesitis-driven disease,²³ in axSpA several evidence indicates bone marrow inflammation is a central early event.^{24,25} Thus, MRI reveals that BME and osteitis frequently precede clinical and radiological enthesitis.²⁴ Hence, the bone marrow provides an immunologically rich milieu where stromal and immune cells propagate inflammation and aberrant bone remodelling.²⁶ Then, communication likely occurs between affected marrow and entheses via cytokines, immune cell trafficking, and anatomical links.²⁶

Genetically, HLA-B27 is the major risk allele, present in up to 80-90% of AS patients,² but the exact pathogenic mechanisms remain unclear. Many other genes related to cytokine signaling, antigen processing, and innate immunity also contribute.²⁷

In terms of immune dysregulation, both arms of the immune system contribute to axSpA pathogenesis. A key pathway implicated is the IL-23/IL-17 axis.²⁸ IL-23 produced by antigenpresenting cells can activate innate lymphoid cells such as type 3 innate lymphoid cells (ILC3s), as well as innate-like T cells including MAIT and $\gamma\delta$ T cells, stimulating them to produce IL-17, IL-22, and other inflammatory mediators. Expanding populations of IL-17-secreting cells drive tissue inflammation and damage.²⁹⁻³² Although adaptive CD4⁺ T_H17 cells also expand in axSpA patients, the relative contribution of innate versus adaptive

sources of IL-17 remains unclear. An imbalance exists between pro-inflammatory T_H17 cells and regulatory T cells that normally maintain self-tolerance. Other relevant cytokines driving inflammation include TNF- α , IL-1, and IL-22. The role of B cells and autoantibodies is still emerging.^{33,34}

Biomechanics contribute through microtrauma and stresses at entheses that could initiate inflammation in susceptible individuals.³⁵ Mechanical instability helps propagate and localize inflammation. Hence, facet joints, SIJ, and spinal entheses endure considerable stresses and are early sites of inflammation.^{36,37}

Environmental triggers like dysbiosis, leaky gut, and infections provide further stimulation. The linkage between gut and joint inflammation supports the gastrointestinal immune environment's contribution.³⁸

In summary our understanding of axSpA pathophysiology has progressed substantially in recent years. The foundation of such evolving perspective is the recognition of the multilayered, intricate interactions underlying disease pathophysiology. Genetic risks, immune dysregulation, biomechanical factors, and environmental triggers collectively propagate aberrant inflammation and tissue damage. Communication likely occurs between affected sites via cytokines, trafficking immune cells, and anatomical connections. Of particular significance is the bone marrow inflammation nowadays considered as an early central event, often preceding clinical signs. MRI bone marrow findings frequently emerge first, providing an immunologically rich nexus where inflammation originates.²⁶

Diagnosis

The diagnosis of axSpA can be challenging due to the lack of a single confirmatory test. AxSpA should be suspected in patients with chronic back pain starting before the age 45 along with signs and symptoms suggestive of SpA. The typical features of inflammatory back pain include insidious onset, improvement with exercise but not rest, pain at night, and morning stiffness lasting over 30 minutes. Other indications for axSpA include presence of HLA-B27, a family history of SpA, elevated C-reactive protein (CRP) levels, extra-articular manifestations (uveitis, psoriasis, inflammatory bowel disease), peripheral arthritis, enthesitis, and good response to non-steroidal anti-inflammatory drugs (NSAIDs).³⁹

Imaging plays a fundamental role in the diagnostics process, with conventional radiography of the SIJ being the recommended first imaging modality in suspected axSpA.⁴⁰ Radiographic sacroiliitis (erosions, sclerosis, joint space widening, ankylosis) confirms a diagnosis, but has low sensitivity in early disease.⁴¹ If radiographs are negative or equivocal. MRI of the SIJ should be performed. MRI can detect BME and osteitis indicating active inflammation. Various structural lesions may also be seen including erosions, sclerosis, fat lesions, and ankylosis. However, there are some limitations with MRI. Bone marrow edema is not entirely specific for axSpA, as it can occur to some degree with mechanical back pain, postpartum, heavy exercise, and even in healthy individuals.^{42,43} Location, extent, and combination with structural lesions may increase specificity. Therefore, MRI interpretation requires experienced readers.^{21,44,45} MRI of the spine has minimal incremental value for diagnosing axSpA when MRI of the SIJ is already performed. However, in patients where SIJ MRI is equivocal or normal, additional spine MRI may increase diagnostic sensitivity by 15-20%.44 On the other hand, it should be taken into account that vertebral corner BME and fat metaplasia also occur in healthy individuals and those with non-specific back pain.⁴⁶ Hence, MRI of both the spine and SIJ is not universally recommended,⁴⁷ although spine MRI can be considered in certain circumstances such as high clinical suspicion despite normal SIJ MRI.⁴⁴ Moreover, spine MRI may predict disease progression since inflammation or fat metaplasia has been traditionally considered to be associated with new syndesmophytes formation,^{48,49} even if new evidence found that vertebral corner inflammation may actually lead to new bone formation, but only in a minority of cases via visible fat deposition.⁵⁰

No serologic markers are confirmatory for axSpA. HLA-B27 positivity has about 90% specificity, but sensitivity around 50%.⁵¹ Elevated CRP supports inflammation but is normal in a relevant proportion of axSpA patients⁵² and can also be seen in other inflammatory circumstances.

Various classification criteria for axSpA have been developed to standardize enrolment in clinical trials and research. However, no universal diagnostic criteria exist. The Assessment of SpondyloArthritis international Society (ASAS) axSpA criteria allow MRI evidence of sacroiliitis to substitute for radiographic damage.¹⁹ It is of great importance to underline that classification criteria might be used and are intended only to recruit a homogeneous group of patients into clinical trials and not to establish a clinical diagnosis. While intended for classification, the ASAS criteria are often used misused clinically, which may lead to some over-diagnosis.⁵³

The diagnosis of axSpA comes from a thorough and multifactorial assessment of the patients which is the combination of clinical, laboratory, genetical and imaging evaluation. There should be a high index of suspicion in the appropriate demographic with suggestive symptoms and signs. Diagnostic evaluation incorporates patient history, physical exam, laboratory tests, and imaging to support the diagnosis, rule out mimics, and assess for poor prognostic factors that may guide therapy.⁵⁴

Treatment

The 2022 ASAS-European Alliance of Associations for Rheumatology (EULAR) recommendations update emphasize a personalized approach to managing axSpA, with treatment tailored to the individual patient. A combination of non-pharmacological and pharmacological treatments is recommended.⁵⁵

For non-pharmacological management, all patients should receive education about axSpA and be encouraged to exercise regularly and stop smoking. Physiotherapy and supervised exercise programs should be considered, particularly for patients who do not exercise independently, as they have proven benefits.⁵⁵

For pharmacological treatment, NSAIDs are recommended as first-line drugs to control pain and stiffness. Continuous NSAID use is preferred if needed to control symptoms, but intermittent 'on-demand' use can be considered if continuous treatment is not required. If NSAID treatment fails, is contraindicated, or poorly tolerated, biological (b) or targeted synthetic (ts) DMARDs should be considered for patients with high disease activity despite conventional treatments. Eligibility criteria include confirmed diagnosis of axSpA: for r-axSpA high disease activity (Ankylosing Spondylitis Disease Activity Score [ASDAS] ≥ 2.1 or Bath Ankylosing Spondylitis Disease Activity Index [BASDAI] ≥ 4) and failure/ contraindications for NSAIDs, while in nr-axSpA in addition to high disease activity and failure/ contraindications for NSAIDs evidence of objective signs of inflammation (elevated CRP and/or positive MRI of the SIJ)⁵⁵ are needed.

According to the international treatment recommendations, it is current practice to start with TNF inhibitors (TNFi) or IL-17 inhibitors due to greater clinical experience and longer safety data. Janus kinase inhibitors (JAKi), like tofacitinib and upadacitinib are newer small molecule tsDMARDs that inhibit intracellular JAK-Signal Transducer and Activator of Transcription (STAT) signaling pathways and have demonstrated efficacy for active axSpA in clinical trials. However, long-term safety data for JAKi in axSpA patients is still limited compared to TNFi and IL-17i. Caution is advised when using JAKi in older patients and those with cardiovascular risk factors, until more safety evidence accumulates. Choice of b/tsDMARD may also be guided by extraarticular manifestations. Anti-TNF monoclonal antibodies are preferred for patients with recurrent uveitis or active IBD, while IL-17i may be preferred for patients with significant skin psoriasis (PsO). If the initial b/tsDMARD fails, switching to another b/tsDMARD should be considered after re-evaluating the diagnosis and comorbidities. On the contrary, if a patient is in sustained remission, tapering of a bDMARD can be considered. At this regard, an increasing number of evidence shows that abruptly withdrawing bDMARDs may lead to a high proportion of flares. On the contrary, tapering was shown to be successful in maintaining treatment response efficiently in a relevant number of patients.55

Radiographic damage and loss of function may require surgical interventions like total hip arthroplasty or spinal osteotomy, even if fortunately, this is not frequently needed anymore. Moreover, potential fractures should be evaluated in patients with sudden worsening symptoms.⁵⁵ In summary, the 2022 ASAS-EULAR recommendations provide an up-to-date, evidence-based guide for optimal management of axSpA, with a focus on individualized treatment approaches.

INNOVATIONS IN THE FIELD OF AXSPA

a. Genetics

Axial spondyloarthritis has a strong genetic component, with heritability over 90%. The major genetic association is with HLA-B27, which accounts for approximately 20-30% of overall disease risk. However, HLA-B27 alone is not sufficient to cause axSpA. In addition to HLA-B27, over 100 non-MHC susceptibility loci have now been identified through genomewide association studies (GWAS).⁵⁶ Each not-HLA variant confers only modest individual risk, reflecting the highly polygenic nature of axSpA. The majority of implicated genes are involved in antigen presentation (ERAP1/2), cytokine signaling (IL23R, IL12B, TYK2), T-cell differentiation (RUNX3, IL7R), and innate immunity (CARD9, TNFRSF1A). These genetic findings implicate altered adaptive and innate immune responses in axSpA pathogenesis.^{27,56,57}

While HLA-B27 is unequivocally the major axSpA genetic factor, its precise molecular role has remained elusive despite extensive research. Proposed mechanisms include altered peptide binding and presentation, induction of endoplasmic reticulum (ER) stress and unfolded protein response, and homodimer formation. Recent T cell repertoire studies have provided some support for the long-debated arthritogenic peptide model.⁵⁸⁻⁶² These have identified expanded clonotypes of CD8⁺ T cells in AS patients that recognize specific HLA-B27-bound self-peptides, suggesting HLA-B27 may aberrantly present certain joint-derived autoantigens to autoreactive T cells. More functional genomics work is still needed to clarify the mechanisms behind most genetic findings in axSpA, including HLA-B27. Resolving these knowledge gaps remains an area of active investigation and is critical to understand axSpA pathogenesis at the molecular level.

Recent genetic studies have revealed intriguing differences between male and female axSpA patients. The association of HLA-B27 appears stronger in men, and certain non-MHC variants like progressive ankylosis protein homolog (ANKH) are associated with r-axSpA specifically in males.^{63,64} Females likely require a higher cumulative genetic burden to develop axSpA, possibly due to X-chromosome effects and protective mechanisms like estrogen. One important observation is that in MRI studies, HLA-B27 associates with more sacroiliac inflammation in axSpA males but not females. In females, factors like obesity and pregnancy history are more relevant to MRI sacroiliitis findings.⁶⁵ This indicates HLA-B27 positivity should be interpreted differently in women with chronic back pain versus men. These sex differences have led to proposals that rather than a classical susceptibility allele, HLA-B27 may act as a modifier influencing disease severity and progression in those who develop axSpA. In this model, HLA-B27 promotes more severe axial inflammation and radiographic changes in males once disease is triggered. In females, axSpA is less HLA-B27 dependent and possibly involves more complex gene-hormone-environment interactions.

The genetic discoveries in axSpA are beginning to enable personalized medicine approaches, although clinical translation remains limited. Polygenic risk scores (PRS) combining hundreds of risk alleles could effectively stratify individuals by genetic disease risk to guide prevention and early intervention.⁶⁶ A polygenic risk score incorporating HLA-B27 and other genetic loci shows excellent predictive power for diagnosing axSpA in European cohorts with a receiver operating curve (ROC) analysis that showed an area under the curve (AUC) of 0.924 (95% CI: 0.920-0.928), superior to HLA-B27 testing alone, the latter with an AUC of 0.869 (95%) CI: 0.865-0.874).67 Pharmacogenomics may eventually allow genetics-tailored treatments, as response to TNFi associates with certain HLA and cytokine genotype variants. However, substantial validation is still required before genetics-based management can be widely implemented. The predictive utility of PRS for prognosis or treatment response is still uncertain. PRS have poor predictive value for general population screening. In addition, most findings derive from populations of European descent, indicating PRS may need customization for diverse ethnic groups.⁶⁷

In summary, ongoing genomic research and translation of genetic findings into clinically applicable tools is critical to achieve the promise of personalized axSpA prevention, diagnosis, prognosis, and treatment based on an individual's genetic makeup.

b. Immunology and serum biomarkers

Innate immune responses likely play the central role initiating inflammation in axSpA. Cells of the innate and innate-like immune systems exhibit functional alterations and accumulate at target tissue sites. Monocytes/macrophages, mast cells, and innate lymphoid cells (ILCs) produce inflammatory cytokines including IL-23, IL-17, TNF- α , and are expanded systemically and locally in axSpA patients.^{31,68,69} Furthermore, neutrophils demonstrate enhanced NETosis.^{70,71} $\gamma\delta$ T cells and mucosal-associated invariant T (MAIT) cells, though technically T cells, exhibit innate-like behaviors and are also implicated as sources of IL-17.^{29,30,72,73}

The contributions of adaptive immunity in disease onset are less defined but likely important in propagating chronic inflammation. Autoreactive TH1 and T_H17 cell populations recognizing joint-derived antigens accumulate in target tissues, which may be driven by molecular mimicry between microbial and self-antigens.^{58,59,74} Though B cells and ectopic lymphoid neogenesis can be detected in tissue lesions, the functional relevance of autoantibodies like anti-CD74 remains uncertain.⁷⁵

The IL-23/IL-17 axis is consistently implicated in early and established axSpA. IL-23 appears important in initial inflammation but becomes less critical once IL-17-secreting innate populations are expanded, which may explain the failure of IL-23 inhibition in treating active axSpA. Thus, ongoing innate production of IL-17 independent of IL-23 stimulation likely sustains chronic inflammation.⁷⁶

Immune function could be influenced and stimulated by several stimuli, each of them known as an etiological factor in axSpA. Therefore, dysbiosis enables translocation of microbial ligands that can trigger innate immune activation both locally and systemically.³⁸ The unfolded protein response and ER stress induced by HLA-B27 misfolding may further stimulate cytokine production in antigen presenting cells.⁶⁰ Finally, biomechanical factors also influence disease initiation and progression, potentially through microdamage sensed by resident innate immune cells.³⁶ In summary, both arms of the immune system make important contributions to axSpA pathogenesis. However, dysregulated innate immunity may play the dominant role in initiating and perpetuating disease; moreover, recent works indicated an involvement of the complement system as well, a cornerstone of the innate immune system.77

The identification of serum biomarkers that improve upon current tools for diagnosing and managing axSpA remains an active area of investigation. Most efforts have focused on analytes reflecting activation of the innate immune pathways implicated in disease. Acute phase reactants like CRP and erythrocyte sedimentation rate (ESR) lack sensitivity and specificity for axSpA. Similarly, cytokines like IL-17, IL-23, TNF- α , and matrix metalloproteinases (MMPs) demonstrate intermittent elevation in patient subgroups but perform inconsistently in diagnosis and monitoring. Calprotectin levels correlate with MRI inflammation in some studies but do not outperform clinical criteria.⁷⁸

Anti-CD74 antibodies exhibit diagnostic potential given their high specificity, but failed validation in certain cohorts. Additional autoantibodies like anti-sclerostin and anti-noggin may have utility in patient subgroups but require larger scale confirmation. Bone turnover markers like sclerostin showed to be reduced in axSpA compared to healthy controls but this is not a specific finding. Circulating collagen fragments, cartilage oligomeric matrix protein (COMP), aggrecan, and other tissue breakdown products appear elevated in-patient serum. Combinations of these biomarkers of cartilage/matrix destruction show promise in predicting MRI and radiographic defined inflammation and damage in preliminary studies. However, they require validation in larger patient cohorts.78

Transcriptomic profiling has identified miRNA signatures that can differentiate axSpA from

chronic back pain. miR-29 is among the most reproducibly dysregulated across studies and influences pathways of relevance to axSpA pathogenesis, although larger studies are needed to validate the clinical utility of miRNA profiles.^{78,79} In summary, currently described serum biomarkers have generally failed to outperform CRP for diagnosing and monitoring axSpA in the clinic. Combinations of tissue breakdown products, autoantibody profiles, microRNAs, and gene scores may eventually provide superior biomarkers to guide management, though extensive further validation is still required.

c. Microbiome

The gut microbiome refers to the vast community of microorganisms inhabiting the gastrointestinal tract. This includes bacteria, viruses, fungi and other microbes living in a complex, interdependent ecosystem. In this regard, dysbiosis refers to an imbalance or alteration of this microbial ecosystem. It is associated with several autoimmune, inflammatory, and metabolic diseases.⁸⁰ In axSpA, particularly, dysbiosis may contribute to disease through various mechanisms:

a) Interaction with HLA-B27

Animal studies of HLA-B27 transgenic rats indicate gut microbes may initiate HLA-B27 misfolding and improper immune reactions that ultimately drive inflammatory responses against joints and spine.^{81,82} For example, germ-free HLA-B27 rats do not develop SpA symptoms, while conventional rats with normal gut microbiota do.⁸³ Specific bacteria like *Klebsiella pneumoniae* showing molecular mimicry to HLA-B27 could be triggering factors.⁸⁴

b) Increased intestinal permeability

In axSpA patients, even without clinically apparent bowel inflammation, microbial imbalance and inflammatory cytokines may impair intestinal barrier integrity. This "leaky gut" enables translocation of bacteria and their products across the intestinal wall, activating immune cells and promoting systemic inflammation. Hence, studies show reduced expression of tight junction proteins and increased intestinal permeability in axSpA patients compared to healthy controls; perhaps, targeting increased gut permeability. States and potential disease modifying strategy.^{84,85}

c) Immune system dysregulation

The gut microbiota interacts extensively with intestinal immune cells, regulating processes like immunoglobuline (Ig)A production, epithelial barrier function, and T-cell differentiation through pattern recognition receptors (PRRs) like toll-like receptors. Dysbiosis in axSpA patients may alter these communications and dysregulate intestinal and systemic immunity. For instance, altered microbiota composition may disrupt the T_H17/ Treg balance, driving pro-inflammatory T_H17 responses.⁸⁶ Segmented filamentous bacteria are potent inducers of T_H17 cells while specific Clostridia strains promote Treg differentiation.^{87,88} Correcting such immune deviations could help restore homeostatic equilibrium.

d) Metabolite changes

The microbiome produces many bioactive metabolites through its metabolic activities. For example, bacterial fermentation of dietary fibers yields short-chain fatty acids (SCFAs) like butyrate which have anti-inflammatory properties.⁸⁹ In axSpA, reduced abundances of SCFA-producing symbionts like *Faecalibacterium prausnitzii* could decrease levels of beneficial metabolites.⁹⁰ Additionally, increased pathobionts or altered microbial gene expression may favor production of detrimental mediators like lipopolysaccharides (LPS), contributing to immune activation. Metabolomic analyses can identify microbiomederived metabolites that drive or suppress inflammation.⁸⁴

Therefore, researchers have made great strides in characterizing the involvement of the gut microbiome in axSpA using advanced genetic sequencing techniques. 16S rRNA gene sequencing of stool samples has revealed altered bacterial diversity and species richness in axSpA patients compared to healthy individuals. This altered diversity indicates an unstable gut ecosystem prone to dysbiosis that lacks resilience against inflammatory triggers.⁹¹ Specific bacterial taxa linked to inflammatory processes appear increased in abundance in the microbiome profiles of axSpA patients. For example, Klebsiella pneumoniae and Proteobacteria, which can trigger HLA-B27 reactions, are often elevated.⁹¹ In contrast, potentially anti-inflammatory bacteria like Faecalibacterium prausnitzii, a major butyrate producer, consistently decline in axSpA

patients.⁸⁹ Reduced levels of beneficial microbes linked to enhanced mucosal barrier function are also commonly observed. The lower amounts of these symbiotic microbes may perpetuate inflammation.^{91,92}

Metagenomic shotgun sequencing provides insights into microbiome functionality by profiling microbial genes and metabolic pathways. In axSpA patients, alterations are seen in genes involved in vitamin biosynthesis, LPS production, and tryptophan metabolism. Of note, tryptophan can be converted to several molecules which are able to shape the function of the immune system and the inflammatory functions, like kynurenines or anti-inflammatory indole derivatives based on the microbial profile.93-95 Importantly, many of these microbiota changes correlate with clinical and inflammatory markers of axSpA. For instance, bacterial dysbiosis associates with levels of CRP, calprotectin, and IL-17. Mucosal inflammation and intestinal lesions are more severe in patients with higher dysbiosis levels. Microbiome parameters also correlate with ASDAS score indicating microbial alterations may reflect axSpA disease activity and pathogenic processes.^{96,97}

In summary, the gut microbiota is complexly involved in axSpA pathogenesis. Ongoing research is unravelling unique microbial signatures in patients and identifying new therapeutic opportunities based on restoring gut homeostasis.

d. Imaging

Imaging plays a critical role in the diagnosis and management of axSpA. Recent technological advances along with standardized image acquisition protocols and validated definitions for positive imaging findings have led to dramatic improvements in axSpA imaging and enhanced diagnostic confidence.

Conventional radiography has been the traditional first-line imaging modality when axSpA is suspected clinically.⁴⁰ However, growing evidence indicates important limitations of radiography in detecting early inflammatory lesions or structural damage compared to advanced cross-sectional imaging now available.^{98,99} Pelvic radiography is widely accessible but imparts radiation exposure. In contrast, multiple studies have demonstrated poor reliability and high interobserver variation in interpreting SIJ radiographs, even among

experienced readers.¹⁰⁰ Compared to MRI or CT, radiography has inferior sensitivity for visualizing the entire spectrum of inflammatory and structural lesions that may develop in the SIJ and spine throughout the disease course of axSpA. Given the clear limitations of radiography and the presence of state-of-the-art alternatives providing unmatched visualization of early axSpA lesions, it may be time to re-assess the role of pelvic radiography as a first-line imaging modality when axSpA is suspected in routine clinical practice.^{101,102}

Magnetic resonance imaging has become an imaging cornerstone of axSpA, offering unparalleled detection of early inflammatory and structural lesions. Recent advances in 3T MRI technology further optimize axSpA evaluation with substantially higher signal-to-noise ratio, improved spatial resolution, and accelerated parallel imaging capabilities compared to conventional 1.5T MRI systems.¹⁰³⁻¹⁰⁵ Consensus definitions for positive MRI findings in axSpA have been proposed through international collaborations like the ASAS MRI group.¹⁰⁶ Recently, this group reported data-driven cut-offs for MRI lesions considered highly suggestive of axSpA after two large-scale reading exercises.¹⁰⁶ Importantly, these cut-offs incorporate both active and structural lesion types. For active lesions, the presence of BME in at least four guadrants of the SIJ or in three consecutive MRI slices demonstrated high specificity for axSpA. The positive predictive value of BME is further increased when erosion or other structural lesions are also visible. Meanwhile, structural lesions including erosions affecting at least three SIJ quadrants or fat metaplasia lesions in five or more quadrants were found to be highly specific for axSpA. Having erosion visible on at least two consecutive MRI slices or fat lesions on at least three consecutive MRI slices was also deemed highly suggestive of axSpA. Fat lesions with a depth over 1 cm were also proposed as a cut-off. These cut-offs reinforce interpreting SIJ MRI based on the collective impact of concomitant inflammatory and structural lesions rather than potentially non-specific findings in isolation. This contextual approach to image assessment enhances diagnostic confidence compared to outdated qualitative paradigms focused predominantly on BME.¹⁰⁶

Beyond conventional MRI, quantitative MRI techniques enable objective, sensitive quantification of inflammation.¹⁰⁷⁻¹⁰⁹ However, substantial work remains to standardize protocols, demonstrate multicenter reproducibility, and validate clinical utility before quantitative MRI is ready for clinical adoption.

Some candidate quantitative MRI methods include T2 mapping, diffusion weighted imaging, and dynamic contrast enhanced MRI. Each of these techniques provides quantitative biomarkers reflecting pathophysiological processes like edema, cellularity, and perfusion. In the future, quantitative MRI has enormous potential to enable sensitive disease monitoring to guide personalized treatment decisions. It could also improve sensitivity for detecting change in clinical trials or observational studies. However, large multicenter trials will be instrumental to validate these techniques across diverse MRI platforms before quantitative MRI can reach its full potential.^{110,111}

While MRI excels at assessing inflammatory lesions, CT remains unsurpassed for visualizing structural bone damage, especially subtle cortical breaks.^{112,113} However, standard CT protocols result in high cumulative radiation exposure, precluding routine use for lifelong monitoring in axSpA patients. Low-dose CT protocols provide an elegant solution through modulating tube current and voltage to substantially reduce radiation dose while maintaining sufficient image quality to assess structural lesions. Noise is controlled through iterative reconstruction algorithms.^{114,115} Early research consistently demonstrates the superiority of low-dose CT protocols compared to radiography for detecting erosions, sclerosis, and syndesmophytes while delivering a similar radiation exposure. Low-dose CT provides an alternate means to evaluate structural damage in cases where MRI is indeterminate or contraindicated.¹¹⁶ However, MRI remains necessary to visualize active inflammation. The precise clinical role for lowdose CT as a supplement to or replacement for MRI or radiography continues to be defined through ongoing studies.

Finally, beyond the traditional visual and qualitative elaboration of imaging data, radiomics involves the high-throughput extraction of quantitative imaging features that can capture tissue heterogeneity and microarchitecture that is not discernible through visual assessment.¹¹⁷⁻¹¹⁹ Radiomics is an emerging technique that may have utility for improving evaluation of axSpA. In radiomics, a large number of quantitative imaging features are extracted from medical images through automated algorithms. Studies have investigated using radiomic analysis of MRI images of the SIJ to identify imaging biomarkers associated with sacroiliitis, SpA diagnosis, and subclassification into axial vs. peripheral subtypes.¹²⁰⁻¹²³ For example, one study extracted over 1,200 texture features from manually segmented SIJ MRI images and identified features that showed significant differences between positive and negative sacroiliitis cases.¹²² A radiomics signature combining multiple features demonstrated good discrimination for diagnosing sacroiliitis with an AUC of 0.82.¹²¹ Another study found certain features differed between axial and peripheral SpA and could distinguish subtypes with excellent accurac.¹²⁰

These preliminary results suggest radiomics can potentially identify imaging biomarkers linked to disease characteristics, activity, and outcomes. This could enable more objective, quantitative evaluation of important MRI features like BME that currently rely on subjective visual assessment. However, there are several limitations. Small sample sizes, lack of independent validation cohorts, and variability in methods across studies make findings exploratory.

There is a need for larger, multicenter studies to validate the reproducibility and added value of radiomic techniques compared to current imaging methods. Extraction of radiomic data requires segmentation of target regions, which can be time-consuming and limit adoption. Automated segmentation methods optimized for SIJ are needed. The complex, multivariate nature of radiomics data also requires specialized biostatistical and machine learning expertise. Despite promising preliminary results, it remains to be determined whether radiomics provides sufficient added diagnostic, prognostic, or monitoring value above current MRI techniques in axial SpA. As methods mature, radiomics may become a useful imaging biomarker for precision medicine approaches, but significant research is still needed to demonstrate clinical utility in SpA.

Axial-spondyloarthritis perspectives

Imaging modality	Role in axial SpA diagnosis and management	Advantages	Limitations	Potential future developments	References
Conventional radiography	Traditional first-line imaging when axSpA is suspected clinically.	Widely accessible	Radiation exposure, limited sensitivity for early inflammatory and structural lesions. High interobserver variation.	Reassess role in routine clinical practice.	40,101
MRI	One-stop shop for detecting early inflammatory and structural lesions.	High sensitivity for both inflammation and structural damage. Ongoing technological advancements (3T MRI).	Requires specialized equipment, expert readers, higher cost compared to radiography.	Quantitative MRI techniques for sensitive inflammation quantification.	103-106
СТ	Unsurpassed for visualizing structural bone damage, including subtle cortical breaks.	Low-dose CT protocols reduce radiation exposure while maintaining image quality.	Radiation exposure, limited for assessing inflammation.	Defining clinical role as a supplement or replacement for MRI or radiography.	112-116
Quantitative MRI	Potential for sensitive quantification of inflammation.	Provides quantitative biomarkers reflecting edema, cellularity, and perfusion.	Standardization, multicenter reproducibility, and clinical validation required.	Enabling personalized treatment and improving sensitivity in clinical trials.	107-111
Radiomics	Emerging technique for improved evaluation.	High-throughput extraction of quantitative imaging features capturing tissue heterogeneity.	Limited by small sample sizes, lack of independent validation, and variability in methods.	Validation in larger multicenter studies, development of automated segmentation methods.	117-121

axSpA: Axial spondyloarthritis; SpA: Spondyloarthritis; MRI: Magnetic resonance imaging; CT: Computed tomography.

In summary, advanced imaging technologies now provide diverse options beyond conventional radiography to assess the myriad lesions occurring in the axial skeleton throughout the course of axSpA. Dedicated SIJ MRI offers unparalleled visualization of early inflammatory and structural damage while low-dose CT provides exquisite detail of cortical breaks. Quantitative MRI shows enormous promise for enabling sensitive quantification of inflammation to guide personalized medicine approaches. Further research and multidisciplinary collaboration will be key to validate these technologies and fully translate their potential to improve patient care into clinical practice (Table 1).

e. Artificial intelligence

Interpretation of axSpA imaging can be challenging due to complex anatomy, variable disease manifestations, and overlap with degenerative changes. There has been increasing interest in using AI and machine learning to improve and automate axSpA imaging analysis.

For conventional radiography, most research has focused on using convolutional neural networks (CNNs) to classify sacroiliitis severity based on the modified New York criteria. Multiple studies have shown CNNs can differentiate normal from definite sacroiliitis (Grade ≥ 2 bilaterally or ≥ 3 unilaterally) with accuracy of 89-97%, sensitivity of 79-91%, and specificity of 79-96%, comparable to rheumatologists. CNNs have also been applied to directly localize SIJ erosions, sclerosis, and ankylosis.¹²⁴

In MRI of the SIJ, common machine learning applications include detecting BME, a hallmark of inflammation.¹²⁵⁻¹²⁸ Various supervised and deep learning approaches have been explored, including thresholding, classical machine learning with hand-crafted features, and CNNs. Reported diagnostic accuracy has been variable, likely related to differences in MRI protocols, gold standards, and class balance across singlecenter studies. However, several CNNs have achieved sensitivity and specificity comparable to experts, with a retrospective multicenter study showing a deep neural network outperforming non-musculoskeletal expert radiologists.¹²⁸ Spatial attention mechanisms, multi-sequence analysis, and clinical data integration have improved model performance. Recent studies show feasibility of automating full SpondyloArthritis Research Consortium of Canada (SPARCC) scoring with CNNs, although reliability in this case remains inferior to human experts.¹²⁹ For spinal MRI, limited research has applied CNNs to detect vertebral corner inflammatory lesions or total inflammatory lesions, but substantial challenges exist in automating full spine analysis given the large search space and lack of robust gold standards.130

Alternative AI applications in axSpA imaging include predicting radiographic progression with CNNs,¹³¹ response to bDMARDs treatments^{132,133} and generating synthetic MRI/CT images. Synthetic CT generated from MRI has recently emerged as a promising technique to improve assessment of structural lesions in axSpA. Deep learning-based algorithms allow reconstruction of CT-like images from specific MRI sequences. Several studies have demonstrated that synthetic CT can visualize SIJ erosions, sclerosis, and ankylosis with greater sensitivity and specificity compared to standard MRI sequences. The improved cortical bone delineation enables more reliable detection of subtle structural lesions that may be overlooked on routine MRI. Enhanced diagnostic performance was confirmed using conventional CT as the reference standard. Synthetic CT imaging may thus expand the utility of MRI for evaluating early structural damage in axSpA, without requiring additional CT radiation exposure.

While AI techniques are being applied to improve imaging-based assessments in axSpA, large language models like GPT-4, LLaMA, Bard or Claude have the potential to act as surrogate patient reported outcome measures (PROs) by generating text summarizing patient symptoms and experience.^{134,135} By analyzing the text from language models, quantitative measures of symptoms could be extracted to track outcomes over time. Potential benefits of using language models as surrogate PROs include reducing patient burden, enabling more frequent tracking of outcomes, and capturing richer qualitative information on patient experience. Challenges include ensuring the language model text accurately reflects patient symptoms, mapping text to quantitative outcomes, and validating performance against traditional PROs.^{134,135}

Despite promising results, adoption of AI techniques remains limited in clinical practice. Key challenges include model generalization across scanners and populations, insufficient training data, variable evaluation frameworks, and lack of clinician trust. Most importantly, large multi-center studies are needed to determine if AI tools improve diagnostic accuracy, enhance workflow efficiency, and benefit patient outcomes compared to standard imaging interpretation. But if AI performance and reliability reaches expert-level, these technologies could expand access to consistent quantitative imaging analysis and objective disease monitoring in axSpA.

CLINICAL IMPLICATIONS

The emerging scientific fields will profoundly transform management of axSpA through enhanced prognostication, quantification, and treatment customization (Figure 2). Genomic medicine has reached an inflection point where polygenic risk scores could soon stratify individuals by genetic disease risk to enable targeted strategies. Advanced imaging modalities now allow direct visualization and sensitive quantification of inflammation in axial joints, paving the way for tightly controlled treat-totarget approaches seeking to achieve remission or low disease activity. The gut microbiome decisive role in shaping immune function suggests future therapies may deliver disease modification by correcting underlying dysbiosis. Together, these advances promise a shift from reactive to predictive, personalized medicine.

Despite remarkable progress, substantial unmet needs persist given the incomplete efficacy and side effects of current biologics; therefore, new therapeutic targets are required and are being investigated nowadays. In this regard, the

pleiotropic cytokine granulocyte macrophagecolony stimulating factor (GM-CSF) promotes multiple inflammatory and osteoproliferative processes in axSpA pathogenesis, hence it has been considered a possible therapeutic target.²² However, phase II results for the GM-CSF inhibitor namilumab were disappointing (NCT03622658). This highlights the ongoing challenge of effectively targeting a single cytokine in a multifactorial disease. In contrast, the dual IL-17A/F inhibitor bimekizumab has now definitively demonstrated efficacy in phase 3 trials for active axSpA,¹³⁶ and therefore added to the therapeutic armamentarium. Looking ahead, the PI3K/Akt/mammalian target of rapamycin (mTOR) pathway represents an appealing target given its dual role regulating aberrant IL-17 responses and pathologic new bone formation.137,138 Preclinical studies indicate PI3Kδ and mTOR inhibition can potentially control both inflammation and osteoproliferation.^{139,140}

Such multifaceted strategies may be essential for enhanced treatment responses.

The potential impact of translating scientific advances into patient benefit remains immense yet unfulfilled. Incorporating polygenic risk scores into predictive algorithms could redefine screening approaches to enable unprecedented early detection in at-risk groups, thereby preventing accumulated damage. MRI inflammation guantification may allow treat-to-target strategies aimed at disease remission or low disease activity, an approach which preliminary evidence suggests correlates with better long-term outcomes. Applying machine learning to standardized imaging data could expand access to quantitative monitoring. However, rigorous comparative effectiveness studies are imperative to validate if emerging tools add value over current standards of care in improving patient-centered outcomes.



Figure 2. New frontiers in axSpA. An overview of emerging areas of research. Key areas highlighted include immunology, genetics, microbiome studies, imaging techniques, and artificial intelligence. Specific examples in each domain are listed. This illustrates the range of innovative techniques being applied to gain new insights into axSpA.

HLA-B27: Human leukocyte antigen B27; ILCs: Innate lymphoid cells; MAIT: Mucosal-associated invariant T; CT: Computed tomography; MRI: Magnetic resonance imaging; LLM: Large language model; axSpA: Axial spondyloarthritis.

INNOVATIONS AND FUTURE DIRECTIONS

Ongoing advances in high-throughput sequencing continue to unravel the intricate geneenvironment interactions underlying pathogenesis, powered by integrative multi-omics approaches defining mechanisms linking the microbiome and immune system. Concurrently, quantitative MRI techniques progress toward clinical adoption for responsive disease monitoring, soon to be augmented by multimodal machine learning tools like hybrid synthetic MRI/CT imaging. Thus, emerging areas will synergize to provide a multidimensional understanding of disease processes from molecular profiling to advanced imaging.

Multiple research gaps remain. Detailed immunophenotyping by single cell sequencing is needed to discern heterogeneity and predict treatment response. The optimal application of microbiome assessment and therapeutics remains uncertain. And crucially, robust validation in large multicenter studies is critical before these tools can be incorporated into updated management guidelines.

Finally, the perspective of patients must remain central in determining unmet needs and developing new technologies. Solving the right problems through co-creation with patients will ensure emerging innovations bring added value to improve outcomes and quality of life. In terms of treatments, new modes of action as well as combination therapies leveraging existing mechanisms are actively being explored to provide more comprehensive disease control. Multitarget approaches may overcome limitations of current monotherapies and lead to superior clinical efficacy. However, careful assessment of safety and real-world effectiveness will be needed as these novel agents and combinations advance through clinical trials.

In conclusion, scientific progress in understanding axSpA pathogenesis has been remarkable, powered by breakthroughs in sequencing, multi-omics profiling, advanced imaging, and machine learning. Collectively, these innovations promise to enable more accurate diagnosis, tightly controlled treatment, and substantially improved patient outcomes. Cross-disciplinary efforts anchored on solving patient needs through co-creation will ultimately determine success in unleashing the benefits of the new fields to reduce the burden of this disease. As we embark on a future illuminated by cutting-edge sequencing, advanced imaging and machine learning, the synergy of these innovations promises a new era in precision medicine, with patient-centric solutions driving transformative outcomes in axSpA.

Data Sharing Statement: The data that support the findings of this study are available from the corresponding author upon reasonable request.

Author Contributions: Conceptualization, review, writing: V.V., S.D.V.; Review, supervision: G.L.; Conceptualization, review, supervision: F.P.

Conflict of Interest: The authors declared no conflicts of interest with respect to the authorship and/or publication of this article.

Funding: The authors received no financial support for the research and/or authorship of this article.

REFERENCES

- 1. Dougados M, Baeten D. Spondyloarthritis. Lancet 2011;377:2127-37. doi: 10.1016/S0140-6736(11)60071-8.
- 2. Sieper J, Poddubnyy D. Axial spondyloarthritis. Lancet 2017;390:73-84. doi: 10.1016/S0140-6736(16)31591-4.
- 3. van der Linden S, Valkenburg HA, Cats A. Evaluation of diagnostic criteria for ankylosing spondylitis. A proposal for modification of the New York criteria. Arthritis Rheum 1984;27:361-8. doi: 10.1002/ art.1780270401.
- 4. Ritchlin C, Adamopoulos IE. Axial spondyloarthritis: New advances in diagnosis and management. BMJ 2021;372:m4447. doi: 10.1136/bmj.m4447.
- 5. López-Medina C, Moltó A. Update on the epidemiology, risk factors, and disease outcomes of axial spondyloarthritis. Best Pract Res Clin Rheumatol 2018;32:241-53. doi: 10.1016/j. berh.2018.10.006.
- Hukuda S, Minami M, Saito T, Mitsui H, Matsui N, Komatsubara Y, et al. Spondyloarthropathies in Japan: Nationwide questionnaire survey performed by the Japan Ankylosing Spondylitis Society. J Rheumatol 2001;28:554-9.
- 7. Mustafa KN, Hammoudeh M, Khan MA. HLA-B27 prevalence in Arab populations and among patients with ankylosing spondylitis. J Rheumatol 2012;39:1675-7. doi: 10.3899/jrheum.120403.

- Andrianakos A, Trontzas P, Christoyannis F, Dantis P, Voudouris C, Georgountzos A, et al. Prevalence of rheumatic diseases in Greece: A cross-sectional population based epidemiological study. The ESORDIG Study. J Rheumatol 2003;30:1589-601.
- Johnsen K, Gran JT, Dale K, Husby G. The prevalence of ankylosing spondylitis among Norwegian Samis (Lapps). J Rheumatol 1992;19:1591-4.
- Dean LE, Jones GT, MacDonald AG, Downham C, Sturrock RD, Macfarlane GJ. Global prevalence of ankylosing spondylitis. Rheumatology (Oxford) 2014;53:650-7. doi: 10.1093/rheumatology/ket387.
- Bohn R, Cooney M, Deodhar A, Curtis JR, Golembesky A. Incidence and prevalence of axial spondyloarthritis: Methodologic challenges and gaps in the literature. Clin Exp Rheumatol 2018;36:263-74.
- 12. Walsh JA, Magrey M. Clinical manifestations and diagnosis of axial spondyloarthritis. J Clin Rheumatol 2021;27:e547-60. doi: 10.1097/ RHU.000000000001575.
- 13. Rudwaleit M, Machado PM, Taieb V, de Peyrecave N, Hoepken B, Gensler LS. Achievement of higher thresholds of clinical responses and lower levels of disease activity is associated with improvements in workplace and household productivity in patients with axial spondyloarthritis. Ther Adv Musculoskelet Dis 2023;15:1759720X231189079. doi: 10.1177/1759720X231189079.
- Frede N, Hiestand S, Endres D, van Elst LT, Finzel S, Chevalier N, et al. Burden of disease and impact on quality of life in chronic back pain - a comparative cross-sectional study of 150 axial spondyloarthritis and 150 orthopedic back pain patients. Front Med (Lausanne) 2023;10:1221087. doi: 10.3389/ fmed.2023.1221087.
- Wilson N, Liu J, Adamjee Q, Di Giorgio S, Steer S, Hutton J, et al. Exploring the emotional impact of axial Spondyloarthritis: A systematic review and thematic synthesis of qualitative studies and a review of social media. BMC Rheumatol 2023;7:26. doi: 10.1186/s41927-023-00351-w.
- Garrido-Cumbrera M, Navarro-Compán V, Bundy C, Mahapatra R, Makri S, Correa-Fernández J, et al. Identifying parameters associated with delayed diagnosis in axial spondyloarthritis: Data from the European map of axial spondyloarthritis. Rheumatology (Oxford) 2022;61:705-12. doi: 10.1093/rheumatology/keab369.
- Redeker I, Callhoff J, Hoffmann F, Haibel H, Sieper J, Zink A, et al. Determinants of diagnostic delay in axial spondyloarthritis: An analysis based on linked claims and patient-reported survey data. Rheumatology (Oxford) 2019;58:1634-8. doi: 10.1093/rheumatology/kez090.
- Deodhar A, Mittal M, Reilly P, Bao Y, Manthena S, Anderson J, et al. Ankylosing spondylitis diagnosis in US patients with back pain: Identifying providers involved and factors associated with rheumatology

referral delay. Clin Rheumatol 2016;35:1769-76. doi: 10.1007/s10067-016-3231-z.

- Rudwaleit M, van der Heijde D, Landewé R, Listing J, Akkoc N, Brandt J, et al. The development of Assessment of SpondyloArthritis international Society classification criteria for axial spondyloarthritis (part II): Validation and final selection. Ann Rheum Dis 2009;68:777-83. doi: 10.1136/ard.2009.108233.
- Sepriano A, Regel A, van der Heijde D, Braun J, Baraliakos X, Landewé R, et al. Efficacy and safety of biological and targeted-synthetic DMARDs: A systematic literature review informing the 2016 update of the ASAS/EULAR recommendations for the management of axial spondyloarthritis. RMD Open 2017;3:e000396. doi: 10.1136/ rmdopen-2016-000396.
- 21. Navarro-Compán V, Ermann J, Poddubnyy D. A glance into the future of diagnosis and treatment of spondyloarthritis. Ther Adv Musculoskelet Dis 2022;14:1759720X221111611. doi: 10.1177/1759720X221111611.
- Del Vescovo S, Venerito V, Iannone C, Lopalco G. Uncovering the underworld of axial spondyloarthritis. Int J Mol Sci 2023;24:6463. doi: 10.3390/ ijms24076463.
- 23. McGonagle D, Lories RJ, Tan AL, Benjamin M. The concept of a "synovio-entheseal complex" and its implications for understanding joint inflammation and damage in psoriatic arthritis and beyond. Arthritis Rheum 2007;56:2482-91. doi: 10.1002/ art.22758.
- Muche B, Bollow M, François RJ, Sieper J, Hamm B, Braun J. Anatomic structures involved in early- and late-stage sacroiliitis in spondylarthritis: A detailed analysis by contrast-enhanced magnetic resonance imaging. Arthritis Rheum 2003;48:1374-84. doi: 10.1002/art.10934.
- Marzo-Ortega H, O'Connor P, Emery P, McGonagle D. Sacroiliac joint biopsies in early sacroiliitis. Rheumatology (Oxford) 2007;46:1210-1. doi: 10.1093/rheumatology/kem098.
- Mauro D, Gandolfo S, Tirri E, Schett G, Maksymowych WP, Ciccia F. The bone marrow side of axial spondyloarthritis. Nat Rev Rheumatol 2023;19:519-32. doi: 10.1038/s41584-023-00986-6.
- Brown MA, Wordsworth BP. Genetics in ankylosing spondylitis - Current state of the art and translation into clinical outcomes. Best Pract Res Clin Rheumatol 2017;31:763-76. doi: 10.1016/j.berh.2018.09.005.
- Taams LS, Steel KJA, Srenathan U, Burns LA, Kirkham BW. IL-17 in the immunopathogenesis of spondyloarthritis. Nat Rev Rheumatol 2018;14:453-66. doi: 10.1038/s41584-018-0044-2.
- 29. Rosine N, Rowe H, Koturan S, Yahia-Cherbal H, Leloup C, Watad A, et al. Characterization of blood mucosal-associated invariant T cells in patients with axial spondyloarthritis and of resident mucosal-associated invariant T cells from the axial

entheses of non-axial spondyloarthritis control patients. Arthritis Rheumatol 2022;74:1786-95. doi: 10.1002/art.42090.

- Toussirot É, Laheurte C, Gaugler B, Gabriel D, Saas P. Increased IL-22- and IL-17A-producing mucosalassociated invariant T cells in the peripheral blood of patients with ankylosing spondylitis. Front Immunol 2018;9:1610. doi: 10.3389/fimmu.2018.01610.
- 31. Ciccia F, Guggino G, Rizzo A, Saieva L, Peralta S, Giardina A, et al. Type 3 innate lymphoid cells producing IL-17 and IL-22 are expanded in the gut, in the peripheral blood, synovial fluid and bone marrow of patients with ankylosing spondylitis. Ann Rheum Dis 2015;74:1739-47. doi: 10.1136/annrheumdis-2014-206323.
- 32. Kenna TJ, Davidson SI, Duan R, Bradbury LA, McFarlane J, Smith M, et al. Enrichment of circulating interleukin-17-secreting interleukin-23 receptor-positive γ/δ T cells in patients with active ankylosing spondylitis. Arthritis Rheum 2012;64:1420-9. doi: 10.1002/art.33507.
- Mauro D, Thomas R, Guggino G, Lories R, Brown MA, Ciccia F. Ankylosing spondylitis: An autoimmune or autoinflammatory disease? Nat Rev Rheumatol 2021;17:387-404. doi: 10.1038/s41584-021-00625-y.
- Li M, Zhou X, Zhou L, Yu Z, Fu L, Yang P. Metaanalysis of changes in the number and proportion of regulatory T cells in patients with ankylosing spondylitis. Biomed Res Int 2020;2020:8709804. doi: 10.1155/2020/8709804.
- 35. Jacques P, McGonagle D. The role of mechanical stress in the pathogenesis of spondyloarthritis and how to combat it. Best Pract Res Clin Rheumatol 2014;28:703-10. doi: 10.1016/j.berh.2014.10.009.
- Cambré I, Gaublomme D, Burssens A, Jacques P, Schryvers N, De Muynck A, et al. Mechanical strain determines the site-specific localization of inflammation and tissue damage in arthritis. Nat Commun 2018;9:4613. doi: 10.1038/s41467-018-06933-4.
- 37. Tinazzi I, McGonagle D, Aydin SZ, Chessa D, Marchetta A, Macchioni P. 'Deep Koebner' phenomenon of the flexor tendon-associated accessory pulleys as a novel factor in tenosynovitis and dactylitis in psoriatic arthritis. Ann Rheum Dis 2018;77:922-5. doi: 10.1136/ annrheumdis-2017-212681.
- 38. Sharip A, Kunz J. Understanding the pathogenesis of spondyloarthritis. Biomolecules 2020;10:1461. doi: 10.3390/biom10101461.
- Navarro-Compán V, Sepriano A, El-Zorkany B, van der Heijde D. Axial spondyloarthritis. Ann Rheum Dis 2021;80:1511-21. doi: 10.1136/ annrheumdis-2021-221035.
- 40. Mandl P, Navarro-Compán V, Terslev L, Aegerter P, van der Heijde D, D'Agostino MA, et al. EULAR recommendations for the use of imaging in the

diagnosis and management of spondyloarthritis in clinical practice. Ann Rheum Dis 2015;74:1327-39. doi: 10.1136/annrheumdis-2014-206971.

- 41. Oostveen J, Prevo R, den Boer J, van de Laar M. Early detection of sacroiliitis on magnetic resonance imaging and subsequent development of sacroiliitis on plain radiography. A prospective, longitudinal study. J Rheumatol 1999;26:1953-8.
- 42. Badr S, Jacques T, Lefebvre G, Boulil Y, Abou Diwan R, Cotten A. Main diagnostic pitfalls in reading the sacroiliac joints on MRI. Diagnostics (Basel) 2021;11:2001. doi: 10.3390/diagnostics11112001.
- 43. Tsoi C, Griffith JF, Lee RKL, Wong PCH, Tam LS. Imaging of sacroiliitis: Current status, limitations and pitfalls. Quant Imaging Med Surg 2019;9:318-35. doi: 10.21037/qims.2018.11.10.
- 44. Aouad K, Maksymowych WP, Baraliakos X, Ziade N. Update of imaging in the diagnosis and management of axial spondyloarthritis. Best Pract Res Clin Rheumatol 2020;34:101628. doi: 10.1016/j. berh.2020.101628.
- 45. De Craemer AS, Łukasik Z, Carron P. Use of imaging in axial spondyloarthritis for diagnosis and assessment of disease remission in the year 2022. Curr Rheumatol Rep 2022;24:383-97. doi: 10.1007/s11926-022-01091-5.
- 46. Weber U, Zubler V, Zhao Z, Lambert RG, Chan SM, Pedersen SJ, et al. Does spinal MRI add incremental diagnostic value to MRI of the sacroiliac joints alone in patients with non-radiographic axial spondyloarthritis? Ann Rheum Dis 2015;74:985-92. doi: 10.1136/annrheumdis-2013-203887.
- 47. Ez-Zaitouni Z, Bakker PA, van Lunteren M, de Hooge M, van den Berg R, Reijnierse M, et al. The yield of a positive MRI of the spine as imaging criterion in the ASAS classification criteria for axial spondyloarthritis: Results from the SPACE and DESIR cohorts. Ann Rheum Dis 2017;76:1731-6. doi: 10.1136/annrheumdis-2017-211486.
- 48. Chiowchanwisawakit P, Lambert RG, Conner-Spady B, Maksymowych WP. Focal fat lesions at vertebral corners on magnetic resonance imaging predict the development of new syndesmophytes in ankylosing spondylitis Arthritis Rheum 2011;63:2215-25. doi: 10.1002/art.30393.
- 49. Machado PM, Baraliakos X, van der Heijde D, Braun J, Landewé R. MRI vertebral corner inflammation followed by fat deposition is the strongest contributor to the development of new bone at the same vertebral corner: A multilevel longitudinal analysis in patients with ankylosing spondylitis. Ann Rheum Dis 2016;75:1486-93. doi: 10.1136/annrheumdis-2015-208011.
- 50. Stal R, Ramiro S, van der Heijde D, van Gaalen FA, Baraliakos X, Machado PM, et al. Do fatty lesions explain the effect of inflammation on new syndesmophytes in patients with radiographic axial spondyloarthritis? Results from the SIAS cohort and

ASSERT trial. RMD Open 2023;9:e003118. doi: 10.1136/rmdopen-2023-003118.

- Komsalova LY, Martínez Salinas MP, Jiménez JFG. Predictive values of inflammatory back pain, positive HLA B27 antigen and acute and chronic magnetic resonance changes in early diagnosis of Spondyloarthritis. A study of 133 patients. PLoS One 2020;15:e0244184. doi: 10.1371/journal. pone.0244184.
- 52. Spoorenberg A, van der Heijde D, de Klerk E, Dougados M, de Vlam K, Mielants H, et al. Relative value of erythrocyte sedimentation rate and C-reactive protein in assessment of disease activity in ankylosing spondylitis. J Rheumatol 1999;26:980-4.
- 53. Proft F, Poddubnyy D. Ankylosing spondylitis and axial spondyloarthritis: Recent insights and impact of new classification criteria. Ther Adv Musculoskelet Dis 2018;10:129-39. doi: 10.1177/1759720X18773726.
- 54. van Gaalen FA, Rudwaleit M. Challenges in the diagnosis of axial spondyloarthritis. Best Pract Res Clin Rheumatol 2023:101871. doi: 10.1016/j. berh.2023.101871.
- 55. Ramiro S, Nikiphorou E, Sepriano A, Ortolan A, Webers C, Baraliakos X, et al. ASAS-EULAR recommendations for the management of axial spondyloarthritis: 2022 update. Ann Rheum Dis 2023;82:19-34. doi: 10.1136/ard-2022-223296.
- Kenyon M, Maguire S, Rueda Pujol A, O'Shea F, McManus R. The genetic backbone of ankylosing spondylitis: How knowledge of genetic susceptibility informs our understanding and management of disease. Rheumatol Int 2022;42:2085-95. doi: 10.1007/s00296-022-05174-5.
- 57. Wordsworth BP, Cohen CJ, Davidson C, Vecellio M. Perspectives on the genetic associations of ankylosing spondylitis. Front Immunol 2021;12:603726. doi: 10.3389/fimmu.2021.603726.
- 58. Faham M, Carlton V, Moorhead M, Zheng J, Klinger M, Pepin F, et al. Discovery of T cell receptor β motifs specific to HLA-B27-positive ankylosing spondylitis by deep repertoire sequence analysis. Arthritis Rheumatol 2017;69:774-84. doi: 10.1002/art.40028.
- 59. Hanson AL, Nel HJ, Bradbury L, Phipps J, Thomas R, Lê Cao KA, et al. Altered repertoire diversity and disease-associated clonal expansions revealed by T cell receptor immunosequencing in ankylosing spondylitis patients. Arthritis Rheumatol 2020;72:1289-302. doi: 10.1002/art.41252.
- Rezaiemanesh A, Mahmoudi M, Amirzargar AA, Vojdanian M, Babaie F, Mahdavi J, et al. Upregulation of unfolded protein response and ER stress-related IL-23 production in M1 macrophages from ankylosing spondylitis patients. Inflammation 2022;45:665-76. doi: 10.1007/s10753-021-01575-z.

- Yu HC, Huang KY, Lu MC, Huang Tseng HY, Liu SQ, Lai NS, et al. HLA-B*27 heavy chain homooligomers promote the cytotoxicity of NK cells via activation of PI3K/AKT signaling. Medicina (Kaunas) 2022;58:1411. doi: 10.3390/medicina58101411.
- Deschler K, Rademacher J, Lacher SM, Huth A, Utzt M, Krebs S, et al. Antigen-specific immune reactions by expanded CD8+ T cell clones from HLA-B*27-positive patients with spondyloarthritis. J Autoimmun 2022;133:102901. doi: 10.1016/j. jaut.2022.102901.
- 63. Rusman T, van Vollenhoven RF, van der Horst-Bruinsma IE. Gender differences in axial spondyloarthritis: Women are not so lucky. Curr Rheumatol Rep 2018;20:35. doi: 10.1007/s11926-018-0744-2.
- 64. Chimenti MS, Perricone C, D'Antonio A, Ferraioli M, Conigliaro P, Triggianese P, et al. Genetics, epigenetics, and gender impact in axialspondyloarthritis susceptibility: An update on genetic polymorphisms and their sex related associations. Front Genet 2021;12:671976. doi: 10.3389/ fgene.2021.671976.
- 65. Braun J, Baraliakos X, Bülow R, Schmidt CO, Richter A. Striking sex differences in magnetic resonance imaging findings in the sacroiliac joints in the population. Arthritis Res Ther 2022;24:29. doi: 10.1186/s13075-021-02712-7.
- Allard-Chamard H, Li Q, Rahman P. Emerging concepts in precision medicine in axial spondyloarthritis. Curr Rheumatol Rep 2023;25:204-12. doi: 10.1007/s11926-023-01113-w.
- 67. Li Z, Wu X, Leo PJ, De Guzman E, Akkoc N, Breban M, et al. Polygenic Risk Scores have high diagnostic capacity in ankylosing spondylitis. Ann Rheum Dis 2021;80:1168-74. doi: 10.1136/ annrheumdis-2020-219446.
- 68. Noordenbos T, Yeremenko N, Gofita I, van de Sande M, Tak PP, Caňete JD, et al. Interleukin-17-positive mast cells contribute to synovial inflammation in spondylarthritis. Arthritis Rheum 2012;64:99-109. doi: 10.1002/art.33396.
- 69. Rosine N, Miceli-Richard C. Innate cells: The alternative source of IL-17 in axial and peripheral spondyloarthritis? Front Immunol 2021;11:553742. doi: 10.3389/fimmu.2020.553742.
- Papagoras C, Chrysanthopoulou A, Mitsios A, Ntinopoulou M, Tsironidou V, Batsali AK, et al. IL-17A expressed on neutrophil extracellular traps promotes mesenchymal stem cell differentiation toward boneforming cells in ankylosing spondylitis. Eur J Immunol 2021;51:930-42. doi: 10.1002/eji.202048878.
- Zambrano-Zaragoza JF, Gutiérrez-Franco J, Durán-Avelar MJ, Vibanco-Pérez N, Ortiz-Martínez L, Ayón-Pérez MF, et al. Neutrophil extracellular traps and inflammatory response: Implications for the immunopathogenesis of ankylosing spondylitis. Int J Rheum Dis 2021;24:426-33. doi: 10.1111/1756-185X.14057.

- 72. Akitsu A, Iwakura Y. Interleukin-17-producing $\gamma\delta$ T ($\gamma\delta$ 17) cells in inflammatory diseases. Immunology 2018;155:418-26. doi: 10.1111/imm.12993.
- Toussirot E, Saas P. MAIT cells: Potent major cellular players in the IL-17 pathway of spondyloarthritis? RMD Open 2018;4:e000821. doi: 10.1136/ rmdopen-2018-000821.
- 74. Zheng M, Zhang X, Zhou Y, Tang J, Han Q, Zhang Y, et al. TCR repertoire and CDR3 motif analyses depict the role of $\alpha\beta$ T cells in Ankylosing spondylitis. EBioMedicine 2019;47:414-26. doi: 10.1016/j.ebiom.2019.07.032.
- 75. Wilbrink R, Spoorenberg A, Verstappen GMPJ, Kroese FGM. B cell involvement in the pathogenesis of ankylosing spondylitis. Int J Mol Sci 2021;22:13325. doi: 10.3390/ijms222413325.
- Navarro-Compán V, Puig L, Vidal S, Ramírez J, Llamas-Velasco M, Fernández-Carballido C, et al. The paradigm of IL-23-independent production of IL-17F and IL-17A and their role in chronic inflammatory diseases. Front Immunol 2023;14:1191782. doi: 10.3389/fimmu.2023.1191782.
- 77. Mistegaard CE, Proft F. The complement system in spondyloarthritis: What do we know. touchREVIEWS in RMD 2022;2:50-6. doi: 10.17925/RMD.2022.1.2.50.
- Reveille JD. Biomarkers in axial spondyloarthritis and low back pain: A comprehensive review. Clin Rheumatol 2022;41:617-34. doi: 10.1007/s10067-021-05968-1.
- 79. Motta F, Carena MC, Selmi C, Vecellio M. MicroRNAs in ankylosing spondylitis: Function, potential and challenges. J Transl Autoimmun 2020;3:100050. doi: 10.1016/j.jtauto.2020.100050.
- Gomes AC, Hoffmann C, Mota JF. The human gut microbiota: Metabolism and perspective in obesity. Gut Microbes 2018;9:308-25. doi: 10.1080/19490976.2018.1465157.
- Hammer RE, Maika SD, Richardson JA, Tang JP, Taurog JD. Spontaneous inflammatory disease in transgenic rats expressing HLA-B27 and human beta 2m: An animal model of HLA-B27-associated human disorders. Cell 1990;63:1099-112. doi: 10.1016/0092-8674(90)90512-d.
- 82. Antoniou AN, Lenart I, Kriston-Vizi J, Iwawaki T, Turmaine M, McHugh K, et al. Salmonella exploits HLA-B27 and host unfolded protein responses to promote intracellular replication. Ann Rheum Dis 2019;78:74-82. doi: 10.1136/annrheumdis-2018-213532.
- Taurog JD, Richardson JA, Croft JT, Simmons WA, Zhou M, Fernández-Sueiro JL, et al. The germfree state prevents development of gut and joint inflammatory disease in HLA-B27 transgenic rats. J Exp Med 1994;180:2359-64. doi: 10.1084/ jem.180.6.2359.
- 84. Song ZY, Yuan D, Zhang SX. Role of the microbiome and its metabolites in ankylosing spondylitis.

Front Immunol 2022;13:1010572. doi: 10.3389/ fimmu.2022.1010572.

- 85. Ciccia F, Guggino G, Rizzo A, Alessandro R, Luchetti MM, Milling S, et al. Dysbiosis and zonulin upregulation alter gut epithelial and vascular barriers in patients with ankylosing spondylitis. Ann Rheum Dis 2017;76:1123-32. doi: 10.1136/ annrheumdis-2016-210000.
- Liu D, Liu B, Lin C, Gu J. Imbalance of peripheral lymphocyte subsets in patients with ankylosing spondylitis: A meta-analysis. Front Immunol 2021;12:696973.doi: 10.3389/fimmu.2021.696973.
- 87. Smith PM, Howitt MR, Panikov N, Michaud M, Gallini CA, Bohlooly-Y M, et al. The microbial metabolites, short-chain fatty acids, regulate colonic Treg cell homeostasis. Science 2013;341:569-73. doi: 10.1126/science.1241165.
- Goto Y, Panea C, Nakato G, Cebula A, Lee C, Diez MG, et al. Segmented filamentous bacteria antigens presented by intestinal dendritic cells drive mucosal Th17 cell differentiation. Immunity 2014;40:594-607. doi: 10.1016/j.immuni.2014.03.005.
- 89. MinHK, NaHS, JhunJ, LeeSY, ChoiSS, ParkGE, etal. Identification of gut dysbiosis in axial spondyloarthritis patients and improvement of experimental ankylosing spondyloarthritis by microbiomederived butyrate with immune-modulating function. Front Immunol 2023;14:1096565. doi: 10.3389/ fimmu.2023.1096565.
- 90. Furusawa Y, Obata Y, Fukuda S, Endo TA, Nakato G, Takahashi D, et al. Commensal microbe-derived butyrate induces the differentiation of colonic regulatory T cells. Nature 2013;504:446-50. doi: 10.1038/nature12721.
- 91. Wang L, Wang Y, Zhang P, Song C, Pan F, Li G, et al. Gut microbiota changes in patients with spondyloarthritis: A systematic review. Semin Arthritis Rheum 2022;52:151925. doi: 10.1016/j. semarthrit.2021.11.002.
- 92. Sorbara MT, Littmann ER, Fontana E, Moody TU, Kohout CE, Gjonbalaj M, et al. Functional and genomic variation between human-derived isolates of lachnospiraceae reveals inter- and intra-species diversity. Cell Host Microbe 2020;28:134-46.e4. doi: 10.1016/j.chom.2020.05.005.
- 93. Scalise G, Ciancio A, Mauro D, Ciccia F. Intestinal microbial metabolites in ankylosing spondylitis. J Clin Med 2021;10:3354. doi: 10.3390/jcm10153354.
- 94. Shen J, Yang L, You K, Chen T, Su Z, Cui Z, et al. Indole-3-acetic acid alters intestinal microbiota and alleviates ankylosing spondylitis in mice. Front Immunol 2022;13:762580. doi: 10.3389/fimmu.2022.762580.
- 95. Berlinberg AJ, Regner EH, Stahly A, Brar A, Reisz JA, Gerich ME, et al. Multi 'omics analysis of intestinal tissue in ankylosing spondylitis identifies alterations in the tryptophan metabolism pathway. Front Immunol 2021;12:587119. doi: 10.3389/fimmu.2021.587119.

- 96. Vallier M, Segurens B, Larsonneur E, Meyer V, Ferreira S, Caloustian C, et al. Characterisation of gut microbiota composition in patients with axial spondyloarthritis and its modulation by TNF inhibitor treatment. RMD Open 2023;9:e002794. doi: 10.1136/rmdopen-2022-002794.
- 97. Sagard J, Olofsson T, Mogard E, Marsal J, Andréasson K, Geijer M, et al. Gut dysbiosis associated with worse disease activity and physical function in axial spondyloarthritis. Arthritis Res Ther 2022;24:42. doi: 10.1186/s13075-022-02733-w.
- 98. Diekhoff T, Eshed I, Radny F, Ziegeler K, Proft F, Greese J, et al. Choose wisely: Imaging for diagnosis of axial spondyloarthritis. Ann Rheum Dis 2022;81:237-42. doi: 10.1136/annrheumdis-2021-220136.
- 99. Protopopov M, Proft F, Wichuk S, Machado PM, Lambert RG, Weber U, et al. Comparing MRI and conventional radiography for the detection of structural changes indicative of axial spondyloarthritis in the ASAS cohort. Rheumatology (Oxford) 2023;62:1631-5. doi: 10.1093/rheumatology/ keac432.
- 100. Protopopov M, Proft F, Sepriano A, Landewé R, van der Heijde D, Maksymowych WP, et al. Radiographic sacroiliitis progression in axial spondyloarthritis: Central reading of 5year followup data from the Assessment of SpondyloArthritis international Society cohort. Rheumatology (Oxford) 2021;60:2478-80. doi: 10.1093/rheumatology/ keab091.
- 101. Eshed I, Diekhoff T, Hermann KGA. Is it time to move on from pelvic radiography as the first-line imaging modality for suspected sacroiliitis? Curr Opin Rheumatol 2023;35:219-25. doi: 10.1097/ BOR.000000000000925.
- 102. Poddubnyy D, Diekhoff T, Baraliakos X, Hermann KGA, Sieper J. Diagnostic evaluation of the sacroiliac joints for axial spondyloarthritis: Should MRI replace radiography? Ann Rheum Dis 2022;81:1486-90. doi: 10.1136/ard-2022-222986.
- 103. Schueller-Weidekamm C, Mascarenhas VV, Sudol-Szopinska I, Boutry N, Plagou A, Klauser A, et al. Imaging and interpretation of axial spondylarthritis: The radiologist's perspective--consensus of the arthritis subcommittee of the ESSR. Semin Musculoskelet Radiol 2014;18:265-79. doi: 10.1055/ s-0034-1375569.
- 104. Ran J, Morelli JN, Xie R, Zhang X, Liang X, Liu X, et al. Role for imaging in spondyloarthritis. Q J Nucl Med Mol Imaging 2017;61:271-82. doi: 10.23736/ S1824-4785.17.02981-8.
- 105. Maksymowych WP, Lambert RG, Østergaard M, Pedersen SJ, Machado PM, Weber U, et al. MRI lesions in the sacroiliac joints of patients with spondyloarthritis: An update of definitions and validation by the ASAS MRI working group. Ann Rheum Dis 2019;78:1550-8. doi: 10.1136/ annrheumdis-2019-215589.

- 106. Maksymowych WP, Lambert RG, Baraliakos X, Weber U, Machado PM, Pedersen SJ, et al. Datadriven definitions for active and structural MRI lesions in the sacroiliac joint in spondyloarthritis and their predictive utility. Rheumatology (Oxford) 2021;60:4778-89. doi: 10.1093/rheumatology/ keab099.
- 107. Keenan KE, Biller JR, Delfino JG, Boss MA, Does MD, Evelhoch JL, et al. Recommendations towards standards for quantitative MRI (qMRI) and outstanding needs. J Magn Reson Imaging 2019;49:e26-39. doi: 10.1002/jmri.26598.
- 108. O'Connor JP, Aboagye EO, Adams JE, Aerts HJ, Barrington SF, Beer AJ, et al. Imaging biomarker roadmap for cancer studies. Nat Rev Clin Oncol 2017;14:169-86. doi: 10.1038/ nrclinonc.2016.162.
- 109. Bloem JL, Reijnierse M, Huizinga TWJ, van der Helm-van Mil AHM. MR signal intensity: Staying on the bright side in MR image interpretation. RMD Open 2018;4:e000728. doi: 10.1136/ rmdopen-2018-000728.
- Martín-Noguerol T, Casado-Verdugo OL, Beltrán LS, Aguilar G, Luna A. Role of advanced MRI techniques for sacroiliitis assessment and quantification. Eur J Radiol 2023;163:110793. doi: 10.1016/j. ejrad.2023.110793.
- 111. Thorley N, Jones A, Ciurtin C, Castelino M, Bainbridge A, Abbasi M, et al. Quantitative magnetic resonance imaging (qMRI) in axial spondyloarthritis. Br J Radiol 2023;96:20220675. doi: 10.1259/ bjr.20220675.
- 112. Lambert RGW, Hermann KGA, Diekhoff T. Low-dose computed tomography for axial spondyloarthritis: Update on use and limitations. Curr Opin Rheumatol 2021;33:326-32. doi: 10.1097/BOR.00000000000803.
- 113. Diekhoff T, Hermann KG, Greese J, Schwenke C, Poddubnyy D, Hamm B, et al. Comparison of MRI with radiography for detecting structural lesions of the sacroiliac joint using CT as standard of reference: Results from the SIMACT study. Ann Rheum Dis 2017;76:1502-8. doi: 10.1136/annrheumdis-2016-210640.
- 114. Chahal BS, Kwan ALC, Dhillon SS, Olubaniyi BO, Jhiangri GS, Neilson MM, et al. Radiation exposure to the sacroiliac joint from low-dose CT compared with radiography. AJR Am J Roentgenol 2018;211:1058-62. doi: 10.2214/AJR.18.19678.
- 115. Willemink MJ, Takx RA, de Jong PA, Budde RP, Bleys RL, Das M, et al. Computed tomography radiation dose reduction: Effect of different iterative reconstruction algorithms on image quality. J Comput Assist Tomogr 2014;38:815-23. doi: 10.1097/RCT.00000000000128.
- 116. Diekhoff T, Hermann KGA, Lambert RG. Future of low-dose computed tomography and dual-energy computed tomography in axial spondyloarthritis.

Curr Rheumatol Rep 2022;24:198-205. doi: 10.1007/s11926-022-01075-5.

- 117. Gillies RJ, Kinahan PE, Hricak H. Radiomics: Images are more than pictures, they are data. Radiology 2016;278:563-77. doi: 10.1148/radiol.2015151169.
- 118. Santos MK, Ferreira Júnior JR, Wada DT, Tenório APM, Barbosa MHN, Marques PMA. Artificial intelligence, machine learning, computer-aided diagnosis, and radiomics: Advances in imaging towards to precision medicine. Radiol Bras 2019;52:387-96. doi: 10.1590/0100-3984.2019.0049.
- 119. Lambin P, Rios-Velazquez E, Leijenaar R, Carvalho S, van Stiphout RG, Granton P, et al. Radiomics: Extracting more information from medical images using advanced feature analysis. Eur J Cancer 2012;48:441-6. doi: 10.1016/j.ejca.2011.11.036.
- 120. Tenório APM, Ferreira-Junior JR, Dalto VF, Faleiros MC, Assad RL, Louzada-Junior P, et al. Radiomic quantification for MRI assessment of sacroiliac joints of patients with spondyloarthritis. J Digit Imaging 2022;35:29-38. doi: 10.1007/ s10278-021-00559-7.
- 121. Tenório APM, Faleiros MC, Junior JRF, Dalto VF, Assad RL, Louzada-Junior P, et al. A study of MRI-based radiomics biomarkers for sacroiliitis and spondyloarthritis. Int J Comput Assist Radiol Surg 2020;15:1737-48. doi: 10.1007/s11548-020-02219-7.
- 122. Ye L, Miao S, Xiao Q, Liu Y, Tang H, Li B, et al. A predictive clinical-radiomics nomogram for diagnosing of axial spondyloarthritis using MRI and clinical risk factors. Rheumatology (Oxford) 2022;61:1440-7. doi: 10.1093/rheumatology/ keab542.
- 123. Zheng M, Miao S, Chen D, Yao F, Xiao Q, Zhu G, et al. Can radiomics replace the SPARCC scoring system in evaluating bone marrow edema of sacroiliac joints in patients with axial spondyloarthritis? Clin Rheumatol 2023;42:1675-82. doi: 10.1007/s10067-023-06543-6.
- 124. Bressem KK, Vahldiek JL, Adams L, Niehues SM, Haibel H, Rodriguez VR, et al. Deep learning for detection of radiographic sacroiliitis: Achieving expert-level performance. Arthritis Res Ther 2021;23:106. doi: 10.1186/s13075-021-02484-0.
- 125. Castro-Zunti R, Park EH, Choi Y, Jin GY, Ko SB. Early detection of ankylosing spondylitis using texture features and statistical machine learning, and deep learning, with some patient age analysis. Comput Med Imaging Graph 2020;82:101718. doi: 10.1016/j.compmedimag.2020.101718.
- 126. Kepp FH, Huber FA, Wurnig MC, Mannil M, Kaniewska M, Guglielmi R, et al. Differentiation of inflammatory from degenerative changes in the sacroiliac joints by machine learning supported texture analysis. Eur J Radiol 2021;140:109755. doi: 10.1016/j.ejrad.2021.109755.
- 127. Roels J, De Craemer AS, Renson T, de Hooge M, Gevaert A, Van Den Berghe T, et al. Machine

learning pipeline for predicting bone marrow edema along the sacroiliac joints on magnetic resonance imaging. Arthritis Rheumatol 2023. doi: 10.1002/ art.42650.

- 128. Bressem KK, Adams LC, Proft F, Hermann KGA, Diekhoff T, Spiller L, et al. Deep learning detects changes indicative of axial spondyloarthritis at MRI of sacroiliac joints. Radiology 2022;305:655-65. doi: 10.1148/radiol.212526.
- 129. Hepburn C, Jones A, Bainbridge A, Ciurtin C, Iglesias JE, Zhang H, et al. Volume of hyperintense inflammation (VHI): A quantitative imaging biomarker of inflammation load in spondyloarthritis, enabled by human-machine cooperation. PLoS One 2023;18:e0284508. doi: 10.1371/journal. pone.0284508.
- 130. Koo BS, Lee JJ, Jung JW, Kang CH, Joo KB, Kim TH, et al. A pilot study on deep learningbased grading of corners of vertebral bodies for assessment of radiographic progression in patients with ankylosing spondylitis. Ther Adv Musculoskelet Dis 2022;14:1759720X221114097. doi: 10.1177/1759720X221114097.
- 131. Baek IW, Jung SM, Park YJ, Park KS, Kim KJ. Quantitative prediction of radiographic progression in patients with axial spondyloarthritis using neural network model in a real-world setting. Arthritis Res Ther 2023;25:65. doi: 10.1186/s13075-023-03050-6.
- 132. Lee S, Kang S, Eun Y, Won HH, Kim H, Lee J, et al. Machine learning-based prediction model for responses of bDMARDs in patients with rheumatoid arthritis and ankylosing spondylitis. Arthritis Res Ther 2021;23:254. doi: 10.1186/s13075-021-02635-3.
- 133. Lee S, Eun Y, Kim H, Cha HS, Koh EM, Lee J. Machine learning to predict early TNF inhibitor users in patients with ankylosing spondylitis. Sci Rep 2020;10:20299. doi: 10.1038/s41598-020-75352-7.
- 134. Venerito V, Bilgin E, Iannone F, Kiraz S. AI am a rheumatologist: A practical primer to large language models for rheumatologists. Rheumatology (Oxford) 2023;62:3256-60. doi: 10.1093/rheumatology/ kead291.
- Venerito V, Puttaswamy D, Iannone F, Gupta L. Large language models and rheumatology: A comparative evaluation. The Lancet Rheumatology 2023;5:e574-8. doi: 10.1016/S2665-9913(23)00216-3.
- 136. van der Heijde D, Deodhar A, Baraliakos X, Brown MA, Dobashi H, Dougados M, et al. Efficacy and safety of bimekizumab in axial spondyloarthritis: Results of two parallel phase 3 randomised controlled trials. Ann Rheum Dis 2023;82:515-26. doi: 10.1136/ard-2022-223595.
- 137. Nagai S, Kurebayashi Y, Koyasu S. Role of PI3K/Akt and mTOR complexes in Th17 cell differentiation. Ann N Y Acad Sci 2013;1280:30-4. doi: 10.1111/ nyas.12059.

- 138. Chen J, Long F. mTOR signaling in skeletal development and disease. Bone Res 2018;6:1. doi: 10.1038/s41413-017-0004-5.
- 139. Chen S, Paveley R, Kraal L, Sritharan L, Stevens E, Dedi N, et al. Selective targeting of PI3K δ suppresses human IL-17-producing T cells and innate-like lymphocytes and may be therapeutic for IL-17-

mediated diseases. J Autoimmun 2020;111:102435. doi: 10.1016/j.jaut.2020.102435.

140. Chen S, van Tok MN, Knaup VL, Kraal L, Pots D, Bartels L, et al. mTOR blockade by rapamycin in spondyloarthritis: Impact on inflammation and new bone formation in vitro and in vivo. Front Immunol 2020;10:2344. doi: 10.3389/fimmu.2019.02344.