

ORIGINAL ARTICLE

Ultrasonographic Measurement of Cartilage Thickness in Patients With Leprosy

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Objectives: This study aims to evaluate femoral cartilage thickness in leprosy patients by using ultrasound.

Patients and methods: Twenty-one patients previously diagnosed with leprosy (11 males and 10 females), and age, sex and body mass index matched 21 controls were enrolled. Control subjects with a history of any other systemic inflammatory disease and/or knee trauma were excluded. All femoral cartilage evaluations were performed by the same physician who was experienced in musculoskeletal ultrasonography using a linear array probe. Measurements for 42 knees of 21 leprosy patients, and 42 knees of 21 healthy subjects were analyzed.

Results: Although patients with leprosy had lower femoral cartilage values than those in the control group, the difference was not statistically significant.

Conclusion: We assume that the non-significantly decreased femoral cartilage thickness in patients with leprosy may be explained by the relative immobilization and biomechanical changes arising from complications.

Key words: Femoral cartilage; leprosy; ultrasonography.

Leprosy is a chronic infectious disease caused by *Mycobacterium leprae*.¹ While the disease primarily affects the skin and/or peripheral nerves,¹ patients may also present with musculoskeletal symptoms such as arthritis, tenosynovitis, enthesitis, pain, weakness, and muscular atrophy involving particularly hands and feet.²⁻⁴ Dermatologists focusing on leprosy should bear in mind that some form of joint involvement may occur in the majority of patients.⁵

Ultrasonography (US), which is increasingly being used as a diagnostic aid in many disorders, has been shown to be a reliable tool for evaluating femoral cartilage.^{6,7} The aim of the current study was to evaluate the femoral cartilage thickness in leprosy patients by using US.

PATIENTS AND METHODS

The study was conducted on out- and inpatients followed at the İstanbul Leprosy Hospital. Ethical approval was obtained from the local ethics committee. All patients were informed about the study protocol and gave a written informed consent.

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Figure 1. Ultrasonographic images (suprapatellar axial view) demonstrating bilateral femoral cartilage measurements of patients with leprosy. R: Right; L: Left; RLC: Right lateral condyle; RIA: Right intercondylar area; RMC: Right medial condyle; LMC: Left medial condyle; LIA: Left intercondylar area; LLC: Left lateral condyle.

Twenty-one patients previously diagnosed with leprosy (11 males and 10 females), and 21 healthy controls (11 males and 10 females) were enrolled. Demographic and clinical features were recorded. Control patients with a history of any other systemic inflammatory diseases such as rheumatoid arthritis, ankylosing spondylitis, spondyloarthritis, systemic lupus erythematosus, gout, and/or knee trauma were excluded.

All femoral cartilage evaluations were performed by the same physician who was experienced in musculoskeletal US using a linear array probe (7-12 MHz, Sonosite M-Turbo 2007-2011, USA). Patients were assessed in supine position with their knees in maximum flexion, and the probe was placed in an axial plane on the suprapatellar area. The femoral cartilage was visualized as a strongly anechoic structure between the bony cortex and the suprapatellar fat. Three mid-point measurements were taken from each knee, at the right lateral condyle (RLC), the right intercondylar area (RIA), the right medial condyle (RMC), the left medial condyle (LMC), the left intercondylar area (LIA), and the left lateral condyle (LLC) (Figure 1).

Statistical analysis

Statistical analyses were performed by using SPSS for Windows version 15.0 (SPSS Inc., Chicago, IL, USA) software program. We expressed variables as mean \pm standard deviation. Since parameters analyzed by Kolmogorov-Smirnov test did not show normal distribution, Mann-Whitney U test and Spearman correlation coefficient analyses were used. A *p* value of <0.05 was considered statistically significant.

RESULTS

Overall, 42 knees of 21 leprosy patients, and 42 knees of 21 healthy subjects were assessed. Three patients had plantar foot ulcers, three had Charcot foot, six had minor lower limb amputations, two had Lisfrank amputation and four had major lower limb amputation. Groups were similar with regards to age, sex, height, weight and body mass index (BMI) (all p>0.05) (Table 1). Mean femoral cartilage thickness values are given in Table 2. Although we observed that patients with leprosy had thinner femoral cartilage values than those in the control group at all measurement sites, the differences did not reach statistical significance (all p>0.05). In addition, correlation analysis between clinical characteristics including age, BMI and disease duration, and cartilage thickness did not reveal significant results (all p>0.05).

Table 1. Demographic fe	atures of	the groups			
	Patient group (n=21)		Control group (n=21)		
	n	Mean±SS	n	Mean±SS	p^*
Age (year)		56.8±13.7		53.3±13.2	0.407
Sex					
Male	11		11		
Female	10		10		
Height (cm)		158.5±9.4		162.7±7.4	0.115
Weight (kg)		69.9± 9.4		67.6±9.2	0.431
Body mass index (kg/m ²)		27.9±3.5		25.5±3.6	0.054
SD: Standard deviation; * The Man	n-Whitney U	test was used to con	npare differ	ences.	

	Patient group (n=21) Control group (n=21)			
	Mean±SS	Mean±SS	р	
Right lateral condyle	0.202±0.21	0.210 ± 0.18	0.386	
Right intercondylar area	0.202±0.23	0.209 ± 0.019	0.331	
Right medial condyle	0.192 ± 0.28	0.199 ± 0.024	0.448	
Left lateral condyle	20.06±0.25	0.211±0.017	0.542	
Left intercondylar area	0.199 ± 0.18	0.208 ± 0.015	0.268	
Left medial condyle	0.191 ± 0.27	0.198 ± 0.025	0.462	

Table 2. Comparison of femoral cartilage thickness measurements (mm)

DISCUSSION

Our results have demonstrated no significant difference in femoral cartilage thickness between patients with leprosy and healthy controls.

Patients with leprosy may present with musculoskeletal symptoms prior to any cutaneous or neurological involvement. While these symptoms may occur at the onset of the disease.⁸ rheumatological manifestations such as softtissue inflammations, arthritis, bone resorption, osteomyelitis, and neuro-osteoarthropathy are usually observed during the reactive inflammatory phase.4,9,10

Peripheral neuropathy associated with motor, sensory, and autonomic function disorders in patients with leprosy has several implications; e.g. contractures of the fingers, muscle weakness, Charcot foot,¹¹⁻¹³ and ulcers leading to minor or major lower limb amputations, all of which may limit mobilization. Patients enrolled in the current study had disorders affecting, in part, their posture and mobilization. Reports on the influence of immobilization on the articular cartilage thickness are controversial.¹⁴ While some reports claimed reduced thickness, some others have reported either increased or unchanged thickness.¹⁵ Healthy distribution of contact stresses in the knee joint is maintained through an intact articular cartilage which in turn depends on a dynamic mechanical loading as well as intermittent hydrostatic pressure.¹⁶ Overall, we consider that the non-significant decrease in the femoral cartilage thickness observed in patients with leprosy may, in part, be explained by the relative immobilization and biomechanical changes arising from leprosy related complications.

On the other hand, this study has several limitations. First, previous reports outlined the influence of hormones (i.e. calcium, vitamin D, estrogen, thyroid hormones) and corticosteroids on the thickness of the cartilage. We did not evaluate these parameters. Second, we did not record the exercise status/mobilization of leprosy patients.

Further researches on the use of US in patients with leprosy, and integrated histological and functional examinations are necessary to demonstrate and thoroughly define the implications of cartilage thickness in this specific group of patients.

Declaration of conflicting interests

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REFERENCES

- 1. Suzuki K, Akama T, Kawashima A, Yoshihara A, Yotsu RR, Ishii N. Current status of leprosy: epidemiology, basic science and clinical perspectives. J Dermatol 2012;39:121-9.
- 2. Slim FJ, Faber WR, Maas M. The role of radiology in nerve function impairment and its musculoskeletal complications in leprosy. Lepr Rev 2009;80:373-87.
- 3. Selvapandian AJ, Sundararaj GD. Infection of the foot and ankle, including leprosy, mycetoma and yaws. In: Jahss M, editor. Disorders of the foot and ankle. Philadelphia: W.B. Saunders Company; 1991. p. 1958-2001.
- 4. Vengadakrishnan K, Saraswat PK, Mathur PC. A study of rheumatological manifestations of leprosy. Indian J Dermatol Venereol Leprol 2004;70:76-8.

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- 5. Chauhan S, Wakhlu A, Agarwal V. Arthritis in leprosy. Rheumatology (Oxford) 2010;49:2237-42.
- Castriota-Scanderbeg A, De Micheli V, Scarale MG, Bonetti MG, Cammisa M. Precision of sonographic measurement of articular cartilage: inter- and intraobserver analysis. Skeletal Radiol 1996;25:545-9.
- Lee CL, Huang MH, Chai CY, Chen CH, Su JY, Tien YC. The validity of in vivo ultrasonographic grading of osteoarthritic femoral condylar cartilage: a comparison with in vitro ultrasonographic and histologic gradings. Osteoarthritis Cartilage 2008;16:352-8.
- Sarkar RN, Phaujdar S, Banerjee S, Siddhanta S, Bhattacharyya K, De D, et al. Musculoskeletal involvement in leprosy. Indian Journal of Rheumatology 2011;6:20-4.
- 9. Pereira HL, Ribeiro SL, Sato EI. Rheumatic manifestations in leprosy. Acta Reumatol Port 2008;33:407-14.
- Cossermelli-Messina W, Festa Neto C, Cossermelli W. Articular inflammatory manifestations in patients with

different forms of leprosy. J Rheumatol 1998;25:111-9.

- Moonot P, Ashwood N, Lockwood D. Orthopaedic complications of leprosy. J Bone Joint Surg [Br] 2005;87:1328-32.
- 12. Van Brakel WH. Peripheral neuropathy in leprosy and its consequences. Lepr Rev 2000;71 Suppl:S146-53.
- Brand PW. The insensitive foot (including leprosy). In: Jahss M, editor. Disorders of the foot and ankle. 2nd ed. Philadelphia: W.B. Saunders; 1991. p. 2170-5.
- Ando A, Suda H, Hagiwara Y, Onoda Y, Chimoto E, Saijo Y, et al. Reversibility of immobilizationinduced articular cartilage degeneration after remobilization in rat knee joints. Tohoku J Exp Med 2011;224:77-85.
- O'Connor KM. Unweighting accelerates tidemark advancement in articular cartilage at the knee joint of rats. J Bone Miner Res 1997;12:580-9.
- Vanwanseele B, Lucchinetti E, Stüssi E. The effects of immobilization on the characteristics of articular cartilage: current concepts and future directions. Osteoarthritis Cartilage 2002;10:408-19.